

# Fixed effects, random effects, repeated measures

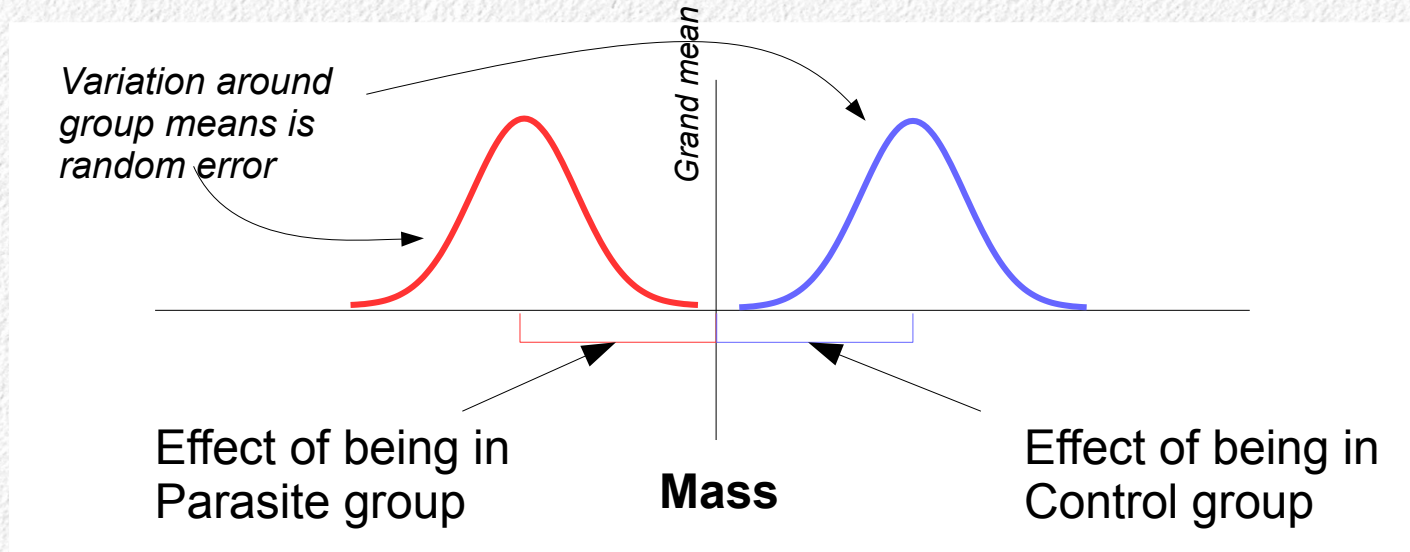
# Fixed effects GLM

- So far, everything we have done is a fixed effects GLM (Model 1)
  - Fixed effects = deterministic, non-random effects of a predictor
  - Can be differences between treatment groups (ANOVA), or numeric relationship between predictor and response (regression)
  - Model coefficients are estimates of fixed effects, which are tested against 0
- Random variation interferes with our measurement of the deterministic effect
- Thus, we estimate the size of a fixed effect, imperfectly, subject to random sampling error

# Example: effects of intestinal parasites on mass of mice

- Question: does parasite treatment affect average mouse weight?
- Experiment:
  - 20 mice randomly assigned to two treatment groups:
    - Parasite larvae (n = 10)
    - Control (n = 10)
  - Each mouse weighed at the end of 1 week
- An F test of  $MS_F/MS_E$  is supposed to tell us whether there is a fixed effect of parasitism
- How, exactly?

# Causes of variation in the data



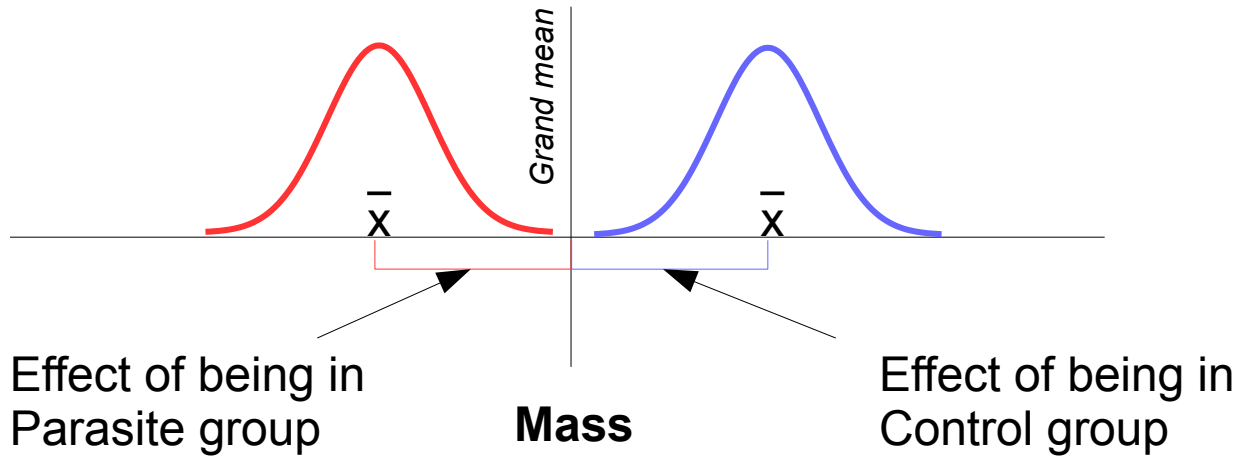
## Fixed effects model:

$$Y = \text{Fixed effect} + \text{error}$$

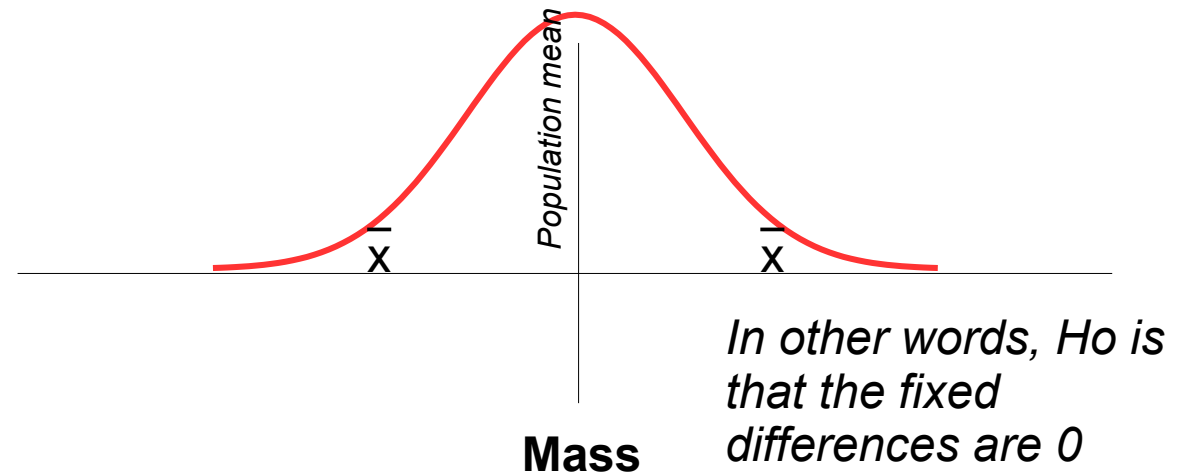
*That is, we treat each mouse's mass as a fixed effect of the parasite group the mouse belongs to plus an unpredictable, random individual contribution*

# What we test with a fixed effects model

Is it this ( $H_A$ )? →



Or this ( $H_0$ )? →



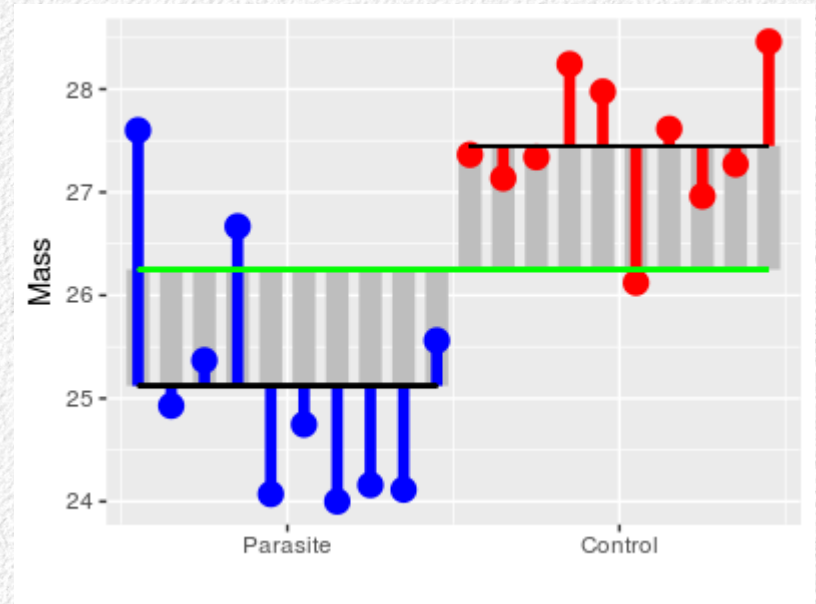
*How does  $MS_F / MS_E$  tell us if there are fixed effects of treatment?*

# Expected mean squares

- **Expected mean squares (E(MS))** = the sources of variation that contribute to a MS calculation
  - Include any fixed effects
  - Include any source of randomness that contributes variation to the estimate of the MS for the term

# Expected mean squares for the fixed effect of parasite treatment

- Effect of treatment is assessed based on group means
- Variation between the group means is due to:
  - Fixed effects of treatment ( $\gamma^2$ )
  - Individual random variation around group means ( $\sigma^2$ )
- Expressed as an *expected* MS for the fixed effect



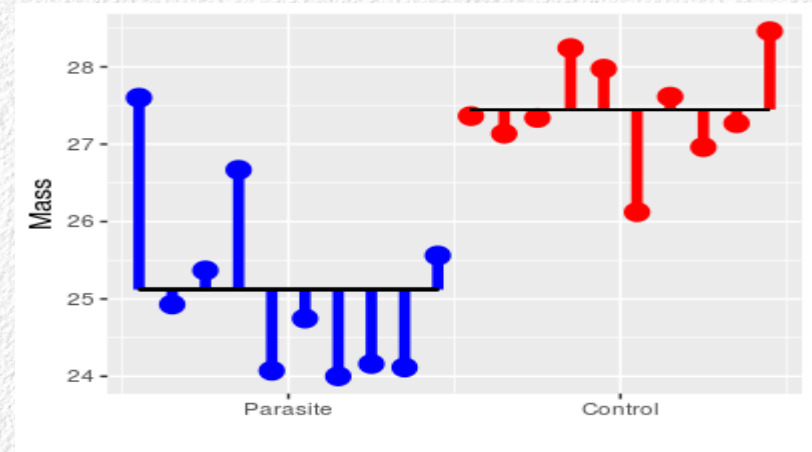
	Df	Sum Sq	Mean Sq	F value	Pr(>F)
treatment	1	27.08	27.078	28.05	4.92e-05
Residuals	18	17.38	0.965		

$$E(MS)_{\text{fixed effect}} = \frac{n \sum \gamma^2}{n-1} + \sigma_E^2$$

Estimated by...

# Expected mean squares for random error

- Variation around group means is all individual, random variation
- Since individual random variation is the only contribution, the expected value is:



	Df	Sum Sq	Mean Sq	F value	Pr(>F)
treatment	1	27.08	27.078	28.05	4.92e-05
Residuals	18	17.38	0.965		

$$E(MS)_{\text{error}} = \sigma_E^2$$

Estimated by...



# Why the F test works

- We use the ratio of two variance estimates ( $MS_F$  and  $MS_E$ ) to test for effects of treatment
- Only difference between  $MS_F$  and  $MS_E$  is that  $MS_F$  contains the fixed effects
- What would  $F_{\text{fixed effect}}$  be if the gammas are 0?

$$E(MS)_{\text{fixed effect}} = \frac{n \sum \gamma^2}{n-1} + \sigma_E^2$$

$$E(MS)_{\text{error}} = \sigma_E^2$$

$$F_{\text{fixed effect}} = \frac{\frac{n \sum \gamma^2}{n-1} + \sigma_E^2}{\sigma_E^2} = \frac{MS_F}{MS_E}$$

# Random effects

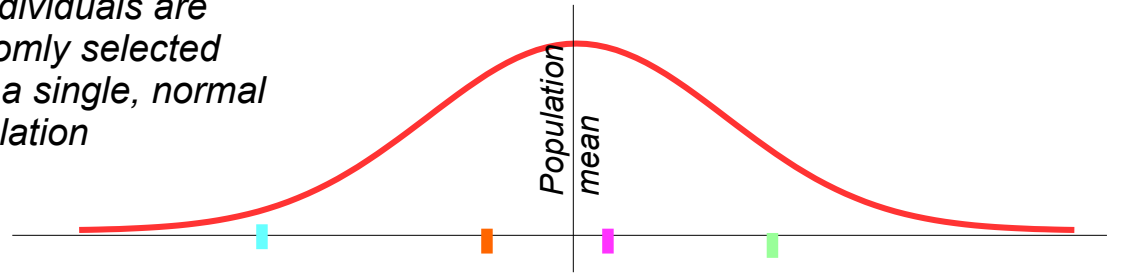
- Different mouse experiment:
  - Randomly select 20 mice
  - Measure each one daily for 11 days
- No fixed effects (no treatment), but there are two different levels of random variation
  - Mice differ randomly in size – a **random effect**
  - Each daily measurement of a mouse can be different – error variation
- A random effect is just another identifiable level of random variation, selected from a distribution
- An ANOVA with only random effects is called Model 2 ANOVA
- SS and MS calculations are done the same way as in Model 1 ANOVA, but the interpretation is different

# Mouse random effect

Two levels of random variation:

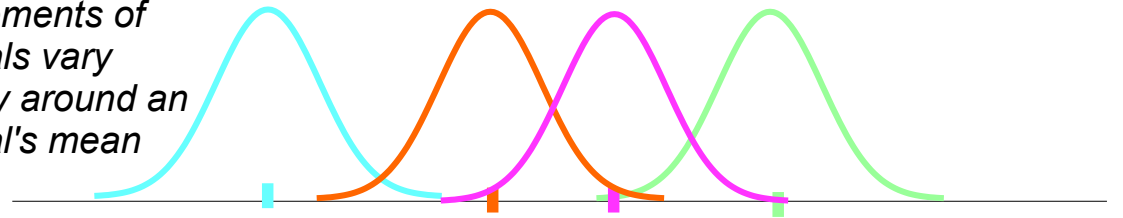
- Among individuals (on average) = mouse random effect
- Among repeated measurements within individuals = error

*All individuals are randomly selected from a single, normal population*



**Mean masses of individuals**

*Repeated measurements of individuals vary randomly around an individual's mean*



**Mass**

**Random effects model:**

$$Y = \text{Random effect} + \text{error}$$

*That is, each measurement is due to the size of the mouse plus random variation in individual daily measurements*

# EMS for random effects

- All variation is random, so use  $\sigma$  as symbol for both levels

- EMS for the mouse level – affected by the variation among individuals, and the random variation among daily measurements

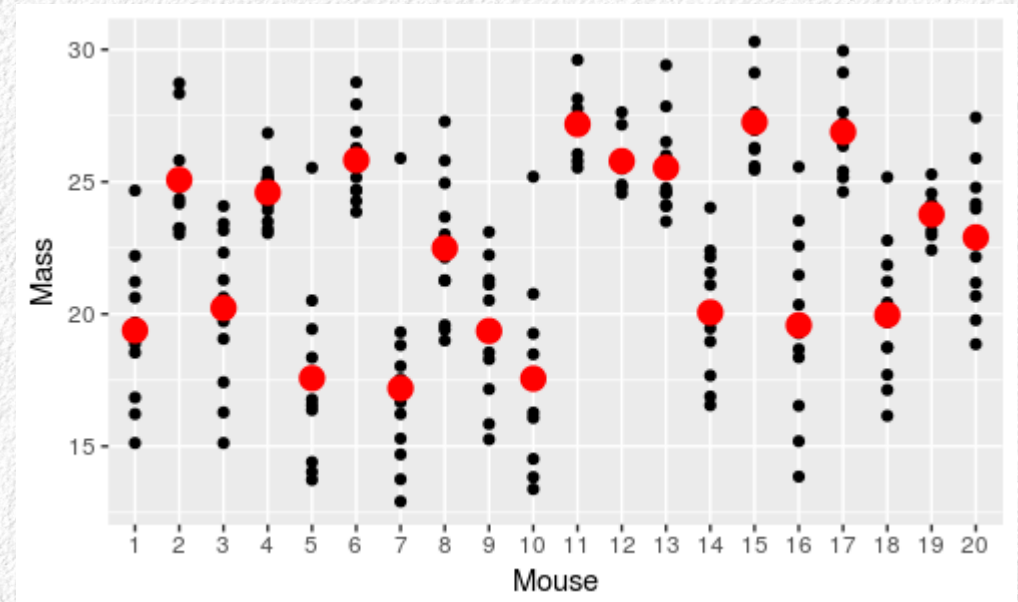
$$E(MS)_{\text{random effect}} = n \sigma_R^2 + \sigma_E^2$$

- EMS for error is still just random variation among daily measurements (i.e. random variation at the lowest level of grouping)

$$E(MS)_{\text{error}} = \sigma_E^2$$

# The data

- Mouse level
  - Mean masses for the mice vary → random effect of mouse
- Error level
  - Daily measurements of each mouse vary around the mouse's mean



# F test for a random effect

- Random effects F test
  - The expected MS tell us to divide the mouse term by the error term
  - Isolates variation among mice
- But, if we run the ANOVA in R...

$$F_{\text{random effect}} = \frac{n\sigma_R^2 + \sigma_E^2}{\sigma_E^2} = \frac{MS_{\text{mouse}}}{MS_E}$$

# Random effects ANOVA table

```
summary(aov(Mass ~ Error(Mouse), data=crmasses))
```

```
Error: mouse
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Residuals	19	2530	133.1		

```
Error: Within
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Residuals	200	1176	5.881		

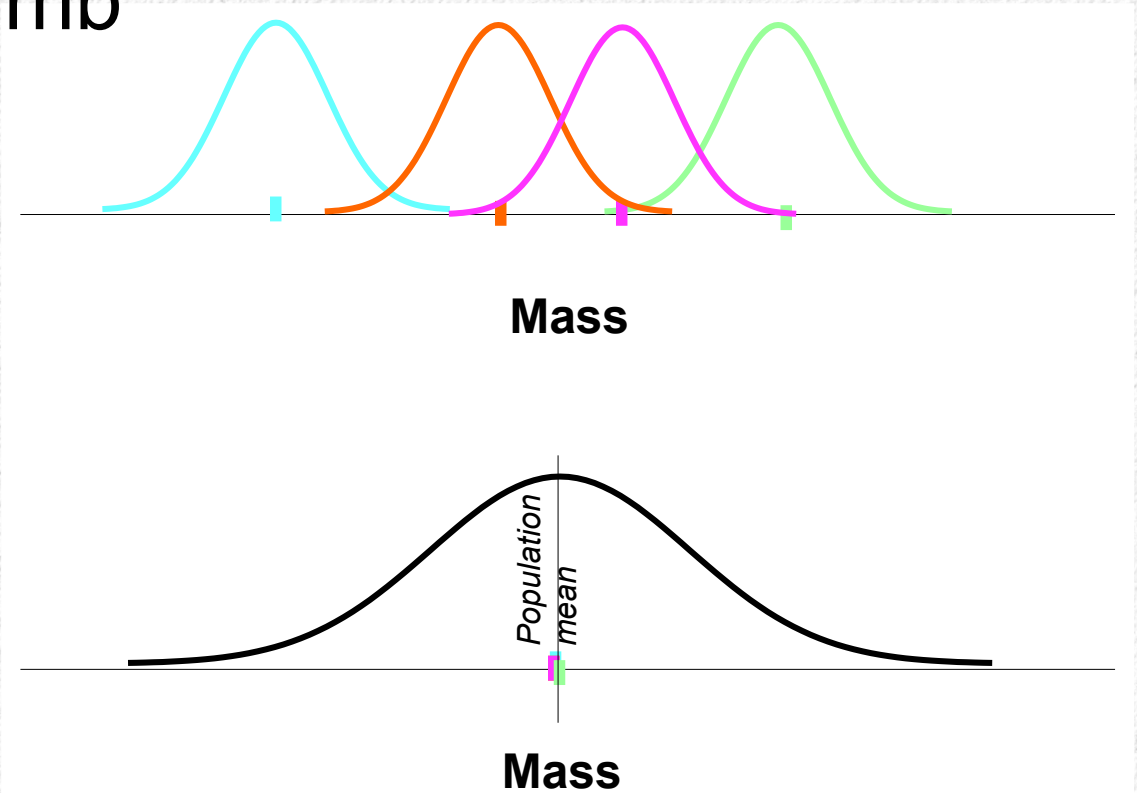
*Need to use the aov() command instead of lm()*

*Error() indicates Mouse is a random effect (an “error strata”, in R lingo)*

*No p-values...why? Because R programmers think p-values for random effects are stupid*

# Why R thinks p-values on random effects are dumb

Is it this ( $H_A$ )?



Or this ( $H_0$ )?



*What are the chances that the null is actually true? Is there any reason to test hypotheses we know are false?*



# Preferred analysis for random effects: variance components

- Treat as an *estimation* problem, not as a hypothesis testing problem
- Estimates of the amount of random variation at each level = **variance components**

- Components are  $\sigma_{mouse}^2$  and  $\sigma_{error}^2$

- R ANOVA table gives us:

$$MS_{mouse} = 133.1 \rightarrow \text{estimate of: } E(MS)_{mouse} = n\sigma_{mouse}^2 + \sigma_{error}^2$$

$$MS_{error} = 5.881 \rightarrow \text{estimate of: } E(MS)_{error} = \sigma_{error}^2$$

- With a little algebra...

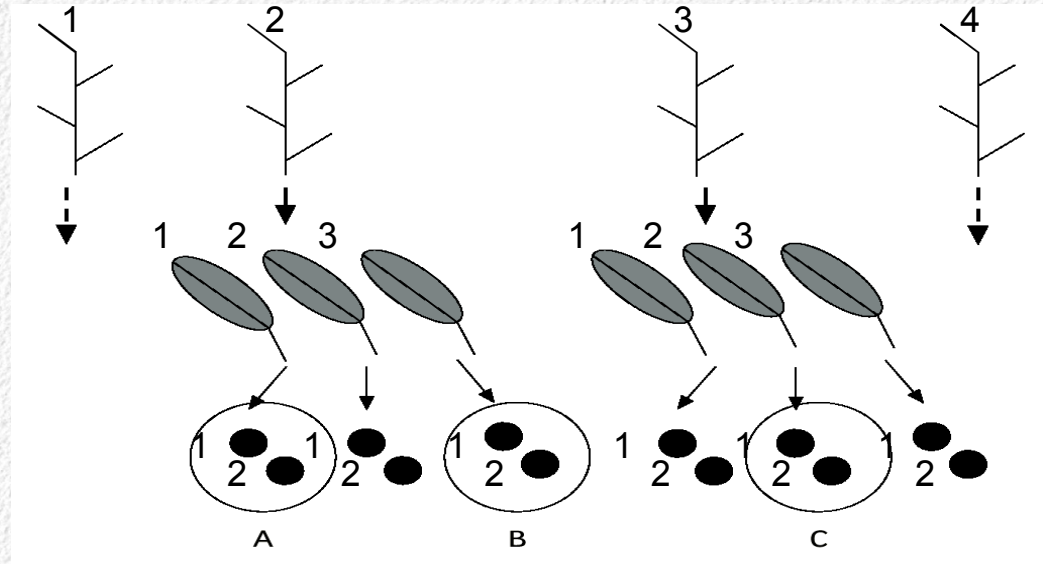
$$\sigma_{mouse}^2 = \frac{MS_{mouse} - MS_{error}}{n} = \frac{133.1 - 5.881}{11} = 11.57$$

$$\sigma_{error}^2 = 5.881$$

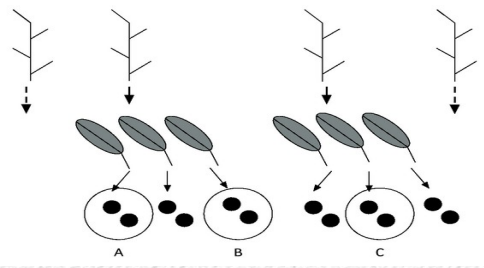
*Conclusion: about twice as much random variation among mice as within*

# Nested designs

- It isn't always possible to cross factors – levels may be nested inside of each other
- Example: measurements of leaf calcium concentration in plants
  - Four plants selected for measurement – levels Plant 1, Plant 2, Plant 3, Plant 4
  - Three leaves from each plant selected – levels Leaf 1, Leaf 2, Leaf 3
  - Two discs cut out of each leaf, measured for calcium concentration
- Leaves are nested within plant
- Discs are nested within leaf
- Random effect of plant, random effect of leaf (disc is error variation)
- Note – numbers are just identifiers (Leaf 1 not a treatment level, just first leaf from each plant)



# EMS for each level



Expected MS for:

Plant  $nm \sigma_{\text{plant}}^2 + n \sigma_{\text{leaf}}^2 + \sigma_E^2$

*Variation of plant means  
around the grand mean*

Leaf  $n \sigma_{\text{leaf}}^2 + \sigma_E^2$

*Variation of leaf means  
around their plant mean*

Error  $\sigma_E^2$

*Variation of discs around  
their leaf means*

# ANOVA table

Call:

```
aov(formula = Conc ~ Error(Plant/Leaf), data = plants)
```

Error: Plant

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Residuals	3	2.724	0.9081		

Error: Plant:Leaf

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Residuals	8	1.27	0.1587		

Error: Within

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Residuals	12	0.5083	0.04236		

# Variance components

$$EMS_{\text{plant}} = nm \sigma_{\text{plant}}^2 + n \sigma_{\text{leaf}}^2 + \sigma_E^2$$

$$MS_{\text{plant}} = 0.9081$$

$$EMS_{\text{leaf}} = n \sigma_{\text{leaf}}^2 + \sigma_E^2$$

$$MS_{\text{leaf}} = 0.1587$$

$$EMS_{\text{error}} = \sigma_E^2$$

$$MS_{\text{error}} = 0.04236$$

$$\sigma_{\text{plant}}^2 = \frac{MS_{\text{plant}} - MS_{\text{leaf}}}{nm} = \frac{0.9081 - 0.1587}{2 \times 3} = 0.1249$$

*Interpretation – greatest variability at plant level, about equal amounts of variability at the leaf and individual disc level*

$$\sigma_{\text{leaf}}^2 = \frac{MS_{\text{leaf}} - MS_{\text{error}}}{n} = \frac{0.1587 - 0.04236}{2} = 0.0582$$

*So? In an experiment, may need lots of plants but not very many leaves per plant, discs per leaf*

# Mixing fixed and random (Model 3 ANOVA)

- What if we:
  - Used 10 parasitized, 10 control mice
  - Weighed them daily for 11 days
- We have a parasite treatment (fixed) and several measurements for individual mice (random) → a **mixed model** (Model 3 ANOVA)
- Problem: which of the two different levels of random variation (mouse, error) should we use to test the parasite effect?
- We can use EMS to tell us which MS is the right denominator for our test of the fixed effect of parasite

# Mixed model EMS

$$E( MS_{par. trt} ) = \frac{n \sum y^2}{n-1} + n \sigma_{mouse}^2 + \sigma_E^2$$

*Effect of parasite subject to random variation between mice and within mice (error)*

$$E( MS_{mouse} ) = n \sigma_{mouse}^2 + \sigma_E^2$$

*EMS for mouse includes both between-mouse and within-mouse*

$$E( MS_{error} ) = \sigma_E^2$$

*EMS for repeated measurements of each mouse (error)*

*What's the appropriate denominator to test the fixed effect of parasite treatment?*

# Mixed effects model in R

aov(Mass ~ Parasite + Error(Mouse))

- Mouse is specified as a random effect with Error(Mouse)
- R knows to calculate a separate error term for Mouse and Residuals
- Also knows that Parasite should be tested over the Mouse term

Error: Mouse

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Parasite	1	2019.3	2019.3	71.24	1.13e-07
Residuals	18	510.2	28.3		

*Using Mouse as the error term is equivalent to averaging by mouse, then using the averages as the data to test for parasite effects*

Error: Within

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Residuals	200	1176	5.881		

*Parasite effect using means for each mouse*

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Parasite	1	183.57	183.57	71.24	1.13e-07
Residuals	18	46.39	2.58		



# Variance components for mouse and residual

Error: Mouse

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Parasite	1	2019.3	2019.3	71.24	1.13e-07
Residuals	18	510.2	28.3		

Error: Within

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Residuals	200	1176	5.881		

$$\sigma_{\text{Mouse}}^2 = \frac{MS_{\text{Mouse}} - MS_{\text{residual}}}{n} = \frac{28.3 - 5.881}{11} = 2.04$$

*So,  $2.04/5.88 = 0.35$ , so random variation between mice is 35% the size of random variation between repeated measurements of the same mouse*

# Problem: repeated measures

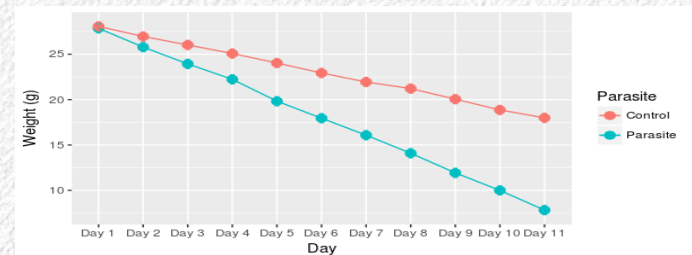
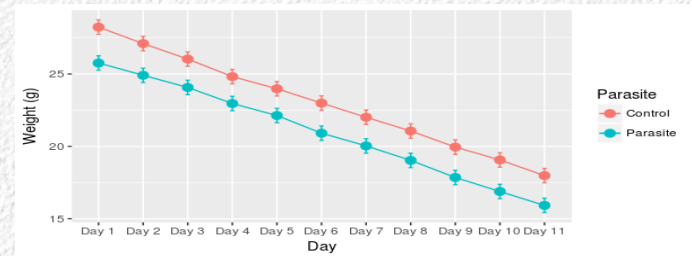
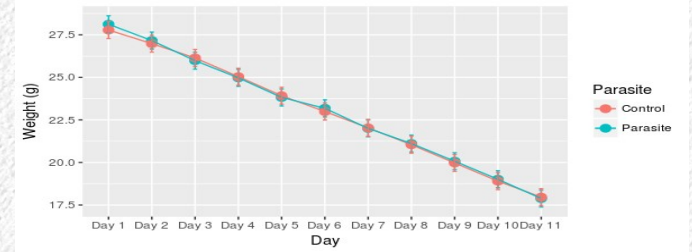
- Repeated measurements of individuals are not independent
  - Pseudoreplication if we use each measurement as a replicate
  - May be patterning in residuals due to change over time
- Using mouse as a random effect only solves the pseudoreplication problem
- We may also be experimentally interested in the change over time
  - When do the treatments become different from one another?
  - How do the patterns of change compare between the treatments?
- Accounting for serially dependent measurements is done with **repeated measures ANOVA**

# The repeated measures design

- Distinguishes **between-subjects** and **within-subjects** effects
  - Between subjects = treatments applied to different subjects (each subject can only be in one group)
  - Within subjects = measurements taken on every subject for every level (i.e. each subject measured every day)
- Parasite treatment is the between-subjects treatment
- Time is the within-subjects treatment

# Main effects and interactions in RMA

- Main effect of parasite indicates that masses differ between parasite and control groups
- Main effect of time indicates change in average mass over time
- Interaction of parasite by time (between x within) indicates that the change over time depends on treatment



# Organization of data

- Each row is an observation of an individual
- Column for Time indicating the time of the measurement (as a factor)
- Mouse column used as a random effect (as a factor)

Mouse	Parasite	Mass	Time
15	Control	30.30	1
8	Control	27.28	1
17	Control	29.95	1
13	Control	29.41	1
12	Control	27.63	1
11	Control	29.61	1
4	Control	26.84	1
19	Control	25.28	1
2	Control	28.34	1
6	Control	28.76	1
20	Treatment	24.17	1
18	Treatment	25.17	1
16	Treatment	25.56	1
14	Treatment	22.15	1
5	Treatment	25.53	1
3	Treatment	23.16	1
1	Treatment	24.67	1
10	Treatment	25.19	1
9	Treatment	21.11	1
7	Treatment	25.89	1
15	Control	29.12	2
8	Control	25.80	2
17	Control	29.12	2

# Repeated measures as a mixed effects model in R

```
aov(Mass ~ Parasite*Time + Error(Mouse))
```

**Error: Mouse**

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
<b>Parasite</b>	<b>1</b>	<b>2019.3</b>	<b>2019.3</b>	<b>71.24</b>	<b>1.13e-07 ***</b>
<b>Residuals</b>	<b>18</b>	<b>510.2</b>	<b>28.3</b>		

*Each mouse measured at each time point, so time only subject to error variation*

**Error: Within**

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
<b>Time</b>	<b>10</b>	<b>937.9</b>	<b>93.79</b>	<b>129.29</b>	<b>&lt;2e-16 ***</b>
<b>Parasite:Time</b>	<b>10</b>	<b>107.8</b>	<b>10.78</b>	<b>14.86</b>	<b>&lt;2e-16 ***</b>
<b>Residuals</b>	<b>180</b>	<b>130.6</b>	<b>0.73</b>		

# An additional assumption of RMA: sphericity

- Sphericity = variances between successive time points are the same
- Sphericity needed for the p-values for the time and time x parasite interactions to be accurate
- If we violate sphericity, we need to:
  - Adjust the p-values (if we don't violate it too badly)
  - Use an approach that doesn't require sphericity called “profile analysis” (if we violate it badly)

# Testing for sphericity, correcting for lack of it

- Tested with the Mauchly test (if  $p < 0.05$ , fail the test)
- Three common corrections – Greenhouse-Geisser, Huynh-Feldt, and lower-bound
  - All three based on “epsilon”, which is 1 when variances are identical across time points, approaches 0 as the variance become increasingly different
  - Epsilon  $> 0.9$  → sphericity is met, p-values in ANOVA table accurate
  - $0.9 > \text{epsilon} > 0.7$  → sphericity is violated, but can use adjusted p-values
  - Epsilon  $< 0.7$  → use profile analysis (multivariate approach)... less powerful than univariate repeated measures when assumptions met, but better option when assumptions not met



# Sphericity failed, use corrected p-values, or profile analysis

## Mauchly Tests for Sphericity

	Test statistic	p-value
time.factor	4.8084e-09	1.033e-30
masses\$Parasite:time.factor	4.8084e-09	1.033e-30

## Greenhouse-Geisser and Huynh-Feldt Corrections for Departure from Sphericity

	GG	eps	Pr(>F[GG])
time.factor	0.21105	<	2.2e-16 ***
masses\$Parasite:time.factor	0.21105	1.265e-05	***

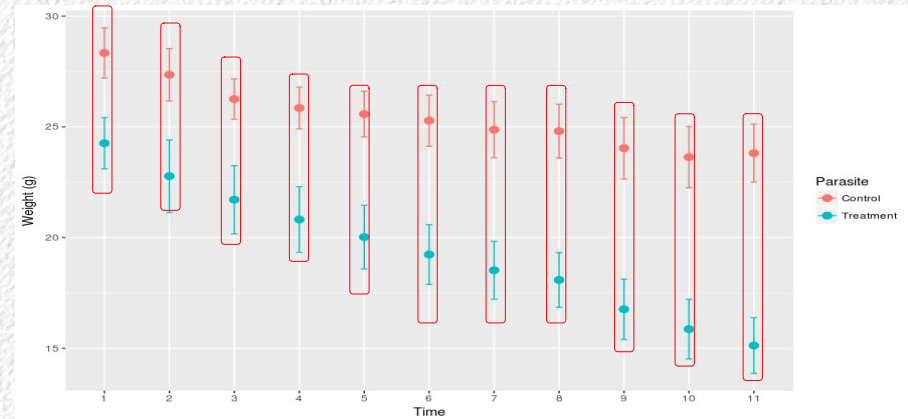
	HF	eps	Pr(>F[HF])
time.factor	0.23978	<	2.2e-16 ***
masses\$Parasite:time.factor	0.23978	3.986e-06	***

# Post-hocs in repeated measures: with no between x within interaction

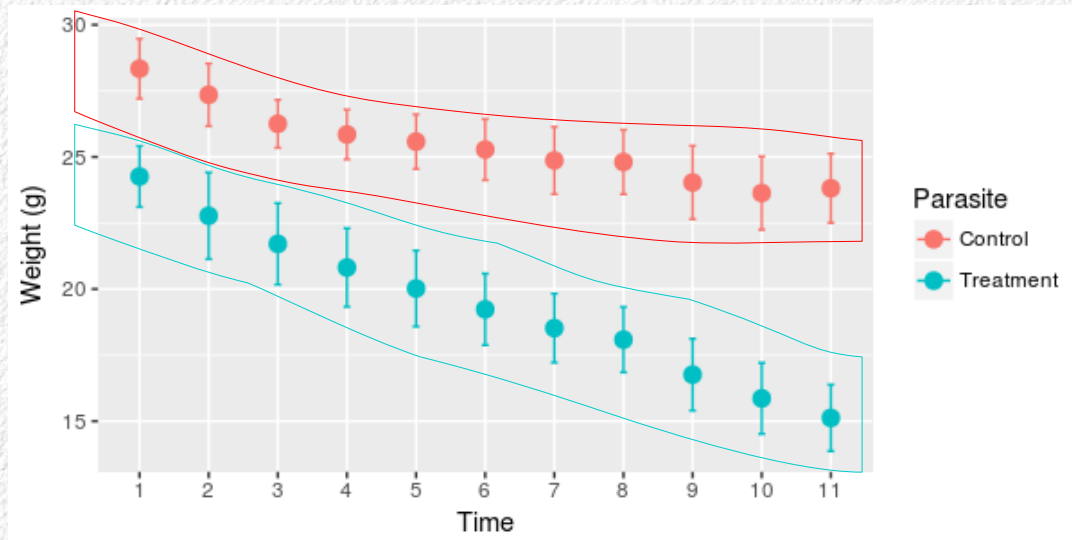
- Within-subjects (time) = compare time points
  - Repeated measurements are paired by individual, should base on **paired** t-tests (why?)
    - Can be all possible pairs of time points
    - Can be only sequential differences
    - Can be initial conditions vs. each subsequent
  - Can use orthogonal polynomials to assess trends over time
- Between-subjects (parasite) – test with Tukey tests

# Post-hocs with an interaction

- Comparing all possible time points between treatments *not* usually desirable
  - 22 combinations of treatment x time
  - 231 pairs of means
- Can assess differences in time trend with orthogonal polynomials
- Can test differences in treatment means at each time point – find the time points at which treatment groups differ



- Can test differences between time points within each treatment group
  - Paired analysis, data subset by parasite group
  - All possible, sequential, against initial



*In either case, would want to use an adjusted alpha level to account for the number of comparisons –  $\alpha/k$*